

Transformation of Major Ginsenosides into Minor Ginsenosides in Ginseng by Pickling in Salted Rice Malt Paste

Manabu Sugimoto^{1*} and Nobutada Murakami²

¹Institute of Plant Science and Resources, Okayama University, 2-20-1 Chuo, Kurashiki, Okayama 710-0046, Japan

²Functional Food Creation Research Institute Co., Ltd., 4422-1 Yoshikawa, Kibichuo, Kaga, Okayama 716-1241, Japan

*Corresponding author

Manabu Sugimoto, Institute of Plant Science and Resources, Okayama University, 2-20-1 Chuo, Kurashiki, Okayama 710-0046, Japan.

Submitted: 25 May 2020; Accepted: 29 May 2020; Published: 05 Jun 2020

Abstract

Increasing the content of minor ginsenosides in ginseng enhances its pharmaceutical activities. For this study, minor ginsenosides F2 and CK were produced in ginseng by pickling in a salted rice malt paste fermented using *Aspergillus kawachii*. The *A. kawachii* excluded β -glucosidase in the culture broth with 125 mU/mL after 16 d cultivation, and hydrolyzed major ginsenoside Rb1 to generate F2 and CK on the pathway Rb1→Rd→F2→CK. Steamed rice was fermented by *A. kawachii* with 14% NaCl to produce a salted rice malt paste as a pickle bed with 165 mU/mL of β -glucosidase. Fresh 1-year-old and 6-year-old ginseng, which had Rb1 and Rd but which had neither F2 nor CK, were pickled in the salted rice malt paste for 28d. The extract showed production of F2 and CK, and changed the ratio of Rb1, Rd2, F2, and CK in ginseng.

Keywords: β -Glucosidase, Compound K (CK), Ginsenoside, Salted Rice Malt Paste, Transformation.

Introduction

Ginseng, the root of *Panax ginseng* C.A. Meyer, is a traditional herbal medicine that has long been used in Asian countries [1]. Ginseng saponins, known as ginsenosides, are major active compounds responsible for the pharmacological effects of ginseng. The more than 50 ginsenosides which have been identified from ginseng comprise a dammarane backbone with several sugar side chains by β -configuration [2, 3]. Ginseng has six major ginsenosides (Rb1, Rb2, Rc, Rd, Re, Rg1), which consist of 80–90% of total ginsenosides [4]. Especially, Rb1 is present in more than 20% of all ginsenosides [5]. Reportedly, major ginsenosides are metabolized to the minor ginsenosides (Rg3, Rh2, F2, CK) by the hydrolyzing activity of intestinal bacteria [5]. Minor ginsenosides show pharmaceutical activities. Among them, particularly CK induces tumor cell apoptosis, inhibition of tumor cell proliferation, attenuation of tumor invasion and metastasis, and anti-inflammatory effects [6–10]. However, the conversion efficiency depends on individual microflora.

Minor ginsenosides have smaller sugar side chains than those of major ginsenosides [11, 12]. Therefore, hydrolysis of the sugar moieties of the major ginsenosides has been developed by microbial or enzymatic biotransformation. Reportedly, minor ginsenosides are produced from purified major ginsenosides or crude extract of ginseng by purified or crude enzyme from microorganisms or cultured microorganisms [12–24]. However, the biotransformation efficiency depends on the substrate specificity of the enzyme. In fact, most microorganisms used for

transformation are not of food production grade. Preparation of the extract of ginseng and enzyme solution costs time and money.

This study revealed that *Aspergillus kawachii*, which is used as koji mold for food fermentation in Japan excretes β -glucosidase to the culture medium well with glucose as the carbon source, thereby hydrolyzing Rb1 to produce F2 and CK through the hydrolyzing pathway of Rb1→Rd→F2→CK. Ginseng pickled in a salted rice malt paste and fermented by *A. kawachii* for 28 d contains F2 and CK and changed the ratio of Rb1, Rd, F2, and CK [25].

Materials and Methods

Materials

One-year-old and 6-year-old ginseng, *Panax ginseng* C.A. Meyer, were obtained from Yuushien Agrifarm Co., Ltd., Shimane, Japan. Ginsenoside Rb1, Rd, and F2 were purchased from LKT Laboratories, Inc. (USA). Also, CK was obtained from MedChem Express LLC (USA). All other chemicals were of analytical and HPLC reagent grade.

Preparation of Culture Broth and Salted Rice Malt Paste

Strains of *Aspergillus oryzae* Cohn var. *oryzae* (NBRC 30113), *A. sojae* Sakaguchi & Yamada (NBRC 4239), *A. kawachii* Kitahara & Yoshida (NBRC 4308), and *A. luchuensis* Inui (NBRC 4314) were obtained from the NITE Biological Resource Center (Tsukuba, Japan). After the strains were inoculated in media consisting of 0.2% NaNO₃, 0.2% KCl, 0.1% KH₂PO₄, 0.1% NH₄NO₃, 0.1% NH₄H₂PO₄, 0.05% MgSO₄, 0.05% yeast extract, and 0.5% glucose (pH 6.0), they were incubated at 26°C with shaking at 120 rpm. The culture solution was filtered through a paper filter. The filtrate was used as a culture broth.

After *A. kawachii* was inoculated into 100 g steamed rice, it was incubated at 28°C for 7 d. The rice with mold was mixed with 1.4 kg steamed rice at 60°C, spread thinly in a Tupperware™ vessel, covered by food wrap, and incubated at 28°C. After 3 d, the rice malt was mixed with 200 g NaCl. After it had been kept in a plastic barrel for 14 d at 23°C in the dark, it was used as a salted rice malt paste.

Preparation of Pickled Ginseng

Pickled ginseng was prepared by soaking 30 g of 1-year-old and 6-year-old ginseng in the salted rice malt paste. The paste was mixed briefly every 2 d and left in the dark at 23°C for 28 d.

Preparation of Ginseng Extract

After 30 g of fresh 1-year-old and 6-year-old ginseng were minced and homogenized in 150 mL of 80% (v/w) ethanol with a Waring blender, the solution was incubated at 80°C. After 14 h, the solution was filtered and evaporated to remove ethanol. The volume of the remaining solution was adjusted to 30 mL with distilled water. It was used as a ginseng extract. The pickled ginseng was washed with water to remove the malt paste. The extract was prepared as described above.

Transformation of Ginsenosides by Culture Broth

The mixture (50 μ L) containing 5 μ L of Rb1 solution (50 μ g) and 45 μ L of culture broth (6 mU β -glucosidase) or the mixture (100 μ L) containing 50 μ L of ginseng extract and 350 μ L of culture broth (44 mU β -glucosidase) was incubated at 37°C for 1, 48, or 96 h. The reaction mixture was extracted with an equal volume of *n*-butanol. Then the *n*-butanol fraction was analyzed using HPLC.

Enzyme Assay

The β -glucosidase activities of the culture solution and salted rice malt paste were assayed using a colorimetric method using

p-nitrophenyl- β -D-glucopyranoside (PNPG) as a substrate [26]. The reaction mixture (1 mL), which consists of 100 mM acetate buffer (pH 5.0), 5 mM PNPG, and culture broth, was incubated at 37°C for 10 min. The reaction was terminated by the addition of 2 mL of 1M sodium bicarbonate. The absorption at 400 nm was measured using a UV spectrophotometer. One unit of β -glucosidase activity was defined as the amount of enzyme liberating 1 μ mol *p*-nitrophenol per min.

HPLC Analysis of Ginsenosides

The ginsenosides were identified using an HPLC analysis system with a column (4.6 \times 250 mm, Cosmosil C₁₈, Nacalai Tesque Inc., Kyoto, Japan) heated at 40°C. The mobile phase consisted of solvent A (water) and solvent B (acetonitrile) with a flow rate of 1.0 mL/min. The initial mobile phase composition of 20% solvent B (0–15 min) changed linearly to 70% (15–30 min). It was then followed by 100% (30–40 min). The effluent was monitored using a UV detector at 203 nm.

Results

β -Glucosidase Activities in Culture Broth and Salted Rice Malt Paste

Aspergillus species of four kinds were cultivated in the medium with glucose as a carbon source. The β -glucosidase activities in the culture broths of *A. oryzae*, *A. sojae*, *A. kawachii*, and *A. luchuensis* were, respectively, 0.50, 0.61, 48, and 6.2 mU/mL after 8 d cultivation, 3.40, 1.90, 97, and 34 mU/mL after 12 d cultivation, and 1.50, 1.10, 125, and 43 mU/mL after 16 d cultivation. The juice of the salted rice malt paste fermented for 14 d was centrifuged at 15,000 \times g for 5 min. The supernatant showed 165 mU/mL β -glucosidase activity.

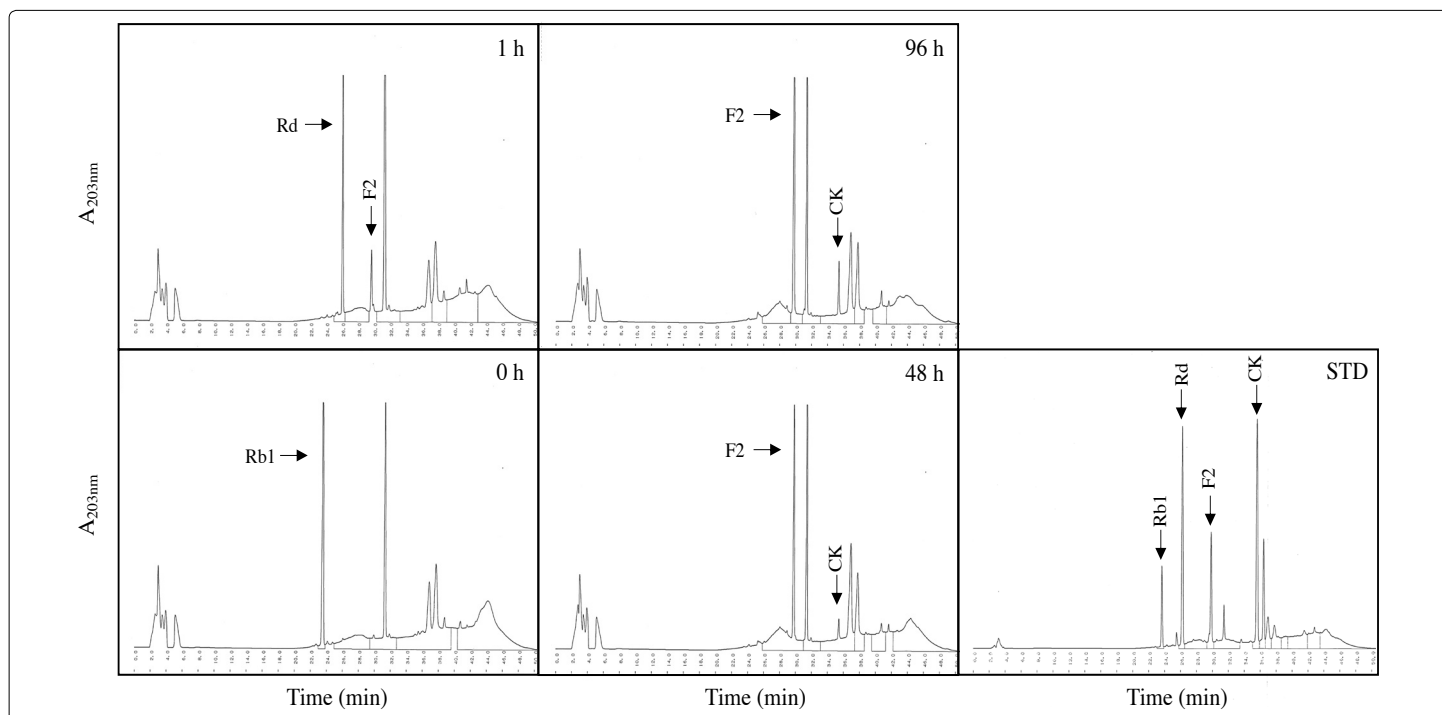


Figure 1: HPLC analysis of the transformation of Rb1 by culture broth of *A. kawachii*

The reaction mixture of Rb1 and culture broth of *A. kawachii* was incubated at 37°C for 1, 48, and 96 h. The reaction mixture was extracted with *n*-butanol; 20 μ L of *n*-butanol fraction was injected. STD, standard mixture containing Rb1, Rd, F2, and CK.

Transformation of Rb1 by Culture Broth

The transformation of Rb1 by the culture broth of *A. kawachii* was analyzed using HPLC (Figure 1). The peak of Rb1 disappeared. Also, Rd and F2 were generated after 1 h, of which the level of Rd was higher than that of F2. After 48 h, the peak of Rd disappeared, the level of F2 increased, and the peak of CK appeared. The level of CK was increased after 96 h. The peak ratios of Rb1, Rd, F2, and CK after 96 h incubation were 0:0:83:17.

Transformation of Ginseng Extract by Culture Broth

Results of HPLC analysis revealed that the extracts 1-year-old and 6-year-old ginseng had Rb1 and Rd, but not F2 and CK. After 96 h incubation of the ginseng extracts with the culture broth of *A. kawachii*, Rb1 and Rd disappeared. Then F2 and CK were generated in both mixtures (Figure 2). The peak ratios of Rb1, Rd, F2, and CK before and after incubation of 1-year-old ginseng extract were 68:32:0:0 and 0:0:87:13, respectively. Those of 6-year-old ginseng extract were 72:28:0:0 and 0:0:83:17, respectively.

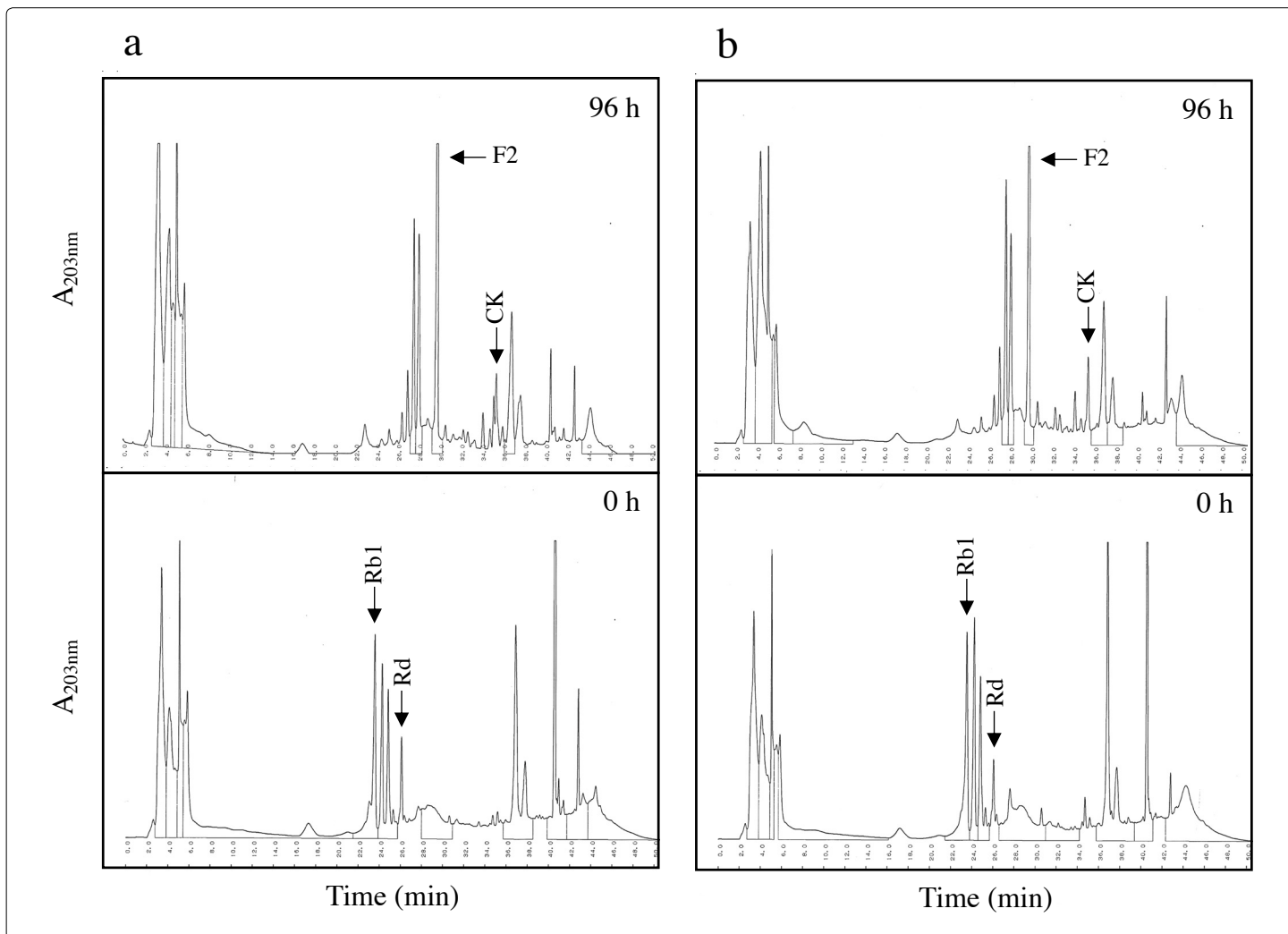


Figure 2: HPLC analysis of the transformation of ginsenosides of extract using culture

Extracts of 1-year-old (a) and 6-year-old (b) ginsengs incubated with the culture broth of *A. kawachii* were incubated at 37°C for 96 h. The reaction mixture was extracted with *n*-butanol; 20 µl of *n*-butanol fraction was injected.

Salt Stability of β-Glucosidase

The salt stability of β-glucosidase in the culture broth of *A. kawachii* was evaluated by incubation with 20% (w/v) NaCl for 60 d at 25°C. The enzyme activities were 117 and 117 mU/mL before and after 60 d incubation, respectively, showing that the enzyme is completely stable after 60 d at 20% NaCl.

Transformation of Ginseng by Salted Rice Malt Paste

The extracts of 1-year-old and 6-year-old ginsengs pickled in the salted rice malt paste for 28 d contained F2 and CK in addition to Rb1 and Rd by HPLC analysis (Figure 3). The ratios of Rb1, Rd, F2, and CK in the extracts of 1-year-old and 6-year-old ginseng roots were, respectively, 40:39:7:14 and 35:45:8:12. The respective amounts of F2 and CK in the 1-year-old ginseng pickled in the salted rice malt paste were 5.0 and 6.0 µg/g; those of pickled 6-year-old ginseng were, respectively, 3.0 and 2.0 µg/g.

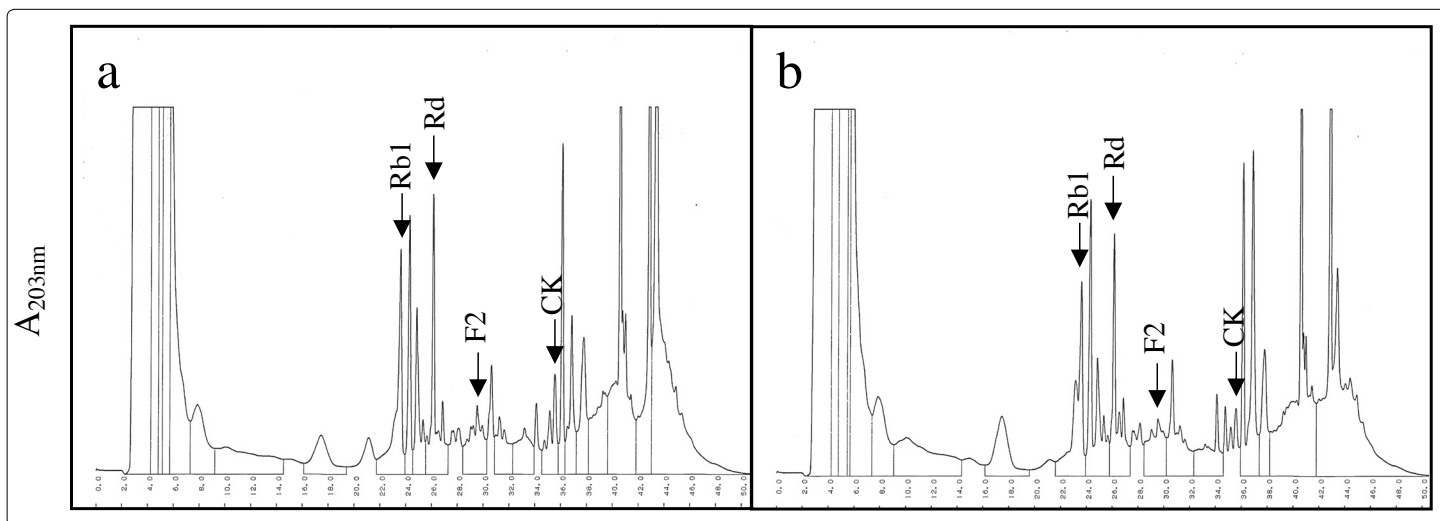


Figure 3: HPLC analysis of extracts of ginsengs pickled in the salted rice mold paste

Ginsenosides of the extracts prepared from 1-year-old and 6-year-old ginsengs pickled in the salted rice mold paste for 28 d were extracted with *n*-butanol; 20 μ l of *n*-butanol fraction was injected: (a) 1-year-old ginseng extract and (b) 6-year-old ginseng extract.

Discussion

Production of minor ginsenosides from major ginsenosides has been approached through the use of microbial and enzymatic reactions. Culture medium of *A. niger*, culture medium of *A. oryzae*, crude enzyme solution from recombinant *Lactococcus lactis*, purified β -glucosidase from *Aspergillus* sp, and purified β -glucosidase from *A. niger* transformed major ginsenosides to minor ginsenosides [21, 22, 27]. These methods require commercial or self-purified major ginsenosides as a substrate, or culture medium or purified enzyme solution. Therefore, they are limited by costs and the working process used for implementation. In addition, a risk exists of mycotoxin contamination because some microorganisms were not edible food microorganisms.

To enhance the contents of minor ginsenosides in ginseng at low labor and cost, we specifically examined Japanese traditional fermentation using “koji mold” and “nukazuke” for transforming ginsenosides in ginseng. Koji mold is used to produce various fermented food products such as sake, soy sauce, and miso. Koji mold produces enzymes such as glycosidases and proteases, which hydrolyze macromolecules such as starches and proteins. Nukazuke is a pickled vegetable in a fermented rice bran paste prepared by mixing rice bran and salt and by subsequent storage in a dark. The pickled vegetables are fermented by lactic acid bacterium and yeast grown in the paste. The major ginsenosides are producible by hydrolysis of sugar moieties from the major ginsenoside using β -glucosidase. Therefore, koji mold is useful as an enzyme source. The nukazuke paste increases the permeability of the pickled vegetables and prevents the growth of toxic microbes. The combinations of the properties inherent in these two methods can change the contents of ginsenosides in ginseng because β -glucosidase excreted from koji mold permeates the ginseng and transforms major ginsenosides into minor ginsenosides while the ginseng is pickled in a rice malt paste fermented by koji mold.

To find the koji mold that excretes β -glucosidase, four typical microorganisms of koji mold, *A. oryzae*, *A. sojae*, *A. kawachii*, and *A. luchuensis*, which have been used respectively to produce sake,

soy sauce, shochu liquor, and awamori liquor, were cultivated in the medium with glucose as a carbon source because steamed rice was used to produce a fermented paste. The *A. kawachii* culture broth showed the highest β -glucosidase activity after 16 d, followed by that of *A. luchuensis*, of which the activity was about one-third that of *A. luchuensis*. The culture broths of *A. oryzae* and *A. sojae* showed maximum activities after 8 d and decreased thereafter. The broth concentrations were, respectively, about 1/37 and 1/66 of the concentration of *A. kawachii*. These results indicate that *A. kawachii* produces extracellular β -glucosidase well in the medium and that it was useful to ferment the steamed rice.

The most abundant ginsenoside in the ginseng is Rb1, which usually accounts for more than 20% of total ginsenosides [5]. Transformation of Rb1 by β -glucosidase from microorganisms has been reported to occur by the hydrolytic pathways Rb1→Rd→F2→CK, Rb1→Rd→Rg3, Rb1→Rd, and Rb1→gypenoside XVII→F2 [1, 12-16, 20-23]. The reaction of culture broth of *A. kawachii* demonstrated that Rb1 disappeared, that Rd and F2 had formed after 1 h, that Rd disappeared, that the level of F2 increased, and that CK had formed by 48 h; moreover, the level of CK had increased after 96 h (Figure 1). The change of ginsenoside contents that is variable in accompaniment to the passage of time suggests the hydrolytic pathway of Rb1→Rd→F2→CK. To evaluate the inhibition of β -glucosidase activity by ginseng contents, the culture broths of *A. kawachii* were incubated with extracts of 1-year-old and 6-year-old ginseng. The Rb1 and Rd in both extracts disappeared; also, F2 and CK were formed after 96 h, indicating that the enzyme can generate F2 and CK (Figure 2). These results suggest that the culture broth of *A. kawachii* can work to produce F2 and CK from Rb1 in ginseng.

High concentrations of salt in the paste are necessary to increase the permeability of the pickled vegetables and to prevent toxic microbe growth. After confirming that β -glucosidase in the culture broth of *A. kawachii* was completely stable after 60 d with 20% NaCl, steamed rice was fermented by *A. kawachii* with 14% NaCl to produce a salted rice malt paste as a pickle bed. The juice of the

paste fermented for 14 d showed 165 mU/mL β -glucosidase activity. Fresh 1-year-old and 6-year-old ginsengs softened when pickled in the paste after 28 d. Whereas the extracts prepared from fresh ginseng showed peaks of Rb1 and Rd but not those of F2 and CK, the peaks of F2 and CK appeared and those of Rb1 and Rd decreased in the extract of the pickled ginseng (Figure 3). These results suggest that the excluded β -glucosidase from *A. kawachii* permeated into the ginseng and transformed Rb1 and Rd into F2 and CK. The contents of F2 and CK in the pickled 1-year-old ginseng were higher than that of the pickled 6-year-old ginseng because the thinner shape and softer peel of 1-year-old ginseng than 6-year-old ginseng probably caused good permeation of β -glucosidase. Taken together, these findings demonstrate that pickling fresh ginseng in the salted rice malt paste is a simple and useful means of producing minor ginsenosides F2 and CK and changing the relative contents of Rb1, Rd, F2, CK in ginseng. Recently CK biosynthesis was achieved by introducing genes encoding enzymes that consist of CK biosynthetic pathway into *Saccharomyces cerevisiae*, *Yarrowia lipolytica*, and *Nicotiana tabacum* [28-31]. These methods are valuable to provide CK for the clinical application. Our pickled ginseng contains F2 and CK in addition to major ginsenosides, which show multifaceted pharmacological activities, while those amounts are small, therefore, it can be used effectively as a resource for dietary supplements, healthy foods, and functional foods [32].

Traditionally, fresh 4-, 5-, and 6-year-old ginseng are used to prepare processed products of ginseng, white ginseng, and red ginseng. White ginseng was produced by drying fresh roots under sunlight. Red ginseng is produced by steaming fresh roots and drying [33]. These processed ginsengs enhance the contents of minor ginsengs and increase their respective pharmaceutical activities [34]. Well-grown and healthy 1-year-old ginseng plants are cultivated to harvest 4-, 5-, and 6-year-old ginseng. After thinning, the remainder of 1-year-old ginseng is discarded or used as a material for eating. Our method of enhancing the minor ginsenosides in ginseng is pickling of ginseng in the salted rice malt paste. Especially, the CK production is expected to elicit biological activities just as it does for white and red ginsengs.

Acknowledgments

We thank Mr. Go Kadowaki, the president of Yuushien Agrifarm Co., Ltd., Shimane, Japan, for supplying 1-year-old and 6-year-old ginseng samples.

References

- Ye L, Zhou CQ, Zhou W, Zhou, P, Chen DF, et al. (2010) Biotransformation of ginsenoside Rb1 to ginsenoside Rd by highly substrate tolerant *Paecilomyces bainier* 229-7. *Bioresource Technology* 101: 7872-7876.
- Jia L, Zhao Y, Liang XJ (2009) Current evaluation of the millennium phytomedicine ginseng (II): collected chemical entities, modern pharmacology, and clinical applications emanated from traditional Chinese medicine. *Current Medical Chemistry* 16: 2924-2942.
- Cheng LQ, Na JR, Bang MH, Kim MK, Yang DC (2008) Conversion of major ginsenoside Rb1 to 20(S)-ginsenoside Rg3 by *Microbacterium* sp. GS514. *Phytochemistry* 69: 218-224.
- Li WK, Gu CG, Zhang HJ, Awang DVC, Fitzloff JF, et al. (2000) Use of high performance liquid chromatography tandem mass spectrometry to distinguish *Panax ginseng* C.A. Meyer (Asian ginseng) and *Panax quinquefolius* L. (American ginseng). *Analytical Chemistry* 72: 5417-5422.
- Ji QC, Harkey MR, Henderson GL, Gershwin ME, Stem JS, et al. (2001) Quantitative determination of ginsenosides by high-performance liquid chromatography-tandem mass spectrometry. *Phytochemical Analysis* 12: 320-326.
- Lee BH, Lee SJ, Hui JH, Lee SY, Sung JH, et al. (1998) In vitro antigenotoxic activity of novel ginseng saponin metabolites formed by intestinal bacteria. *Planta Medica* 64: 500-503.
- Bae EA, Choo MK, Park SY, Shin SY, Kim DH (2002) Metabolism of ginsenoside Rc by human intestinal bacteria and its related antiallergic activity. *Biological Pharmaceutical Bulletin* 25: 743-747.
- Nag SA, Qin JJ, Wang W, Wang MH, Wang H, et al. (2012) Ginsenosides as anticancer agents: In vitro and in vivo activities, structure activity relationships, and molecular mechanisms of action. *Frontiers Pharmacology* 3: 25.
- Li HU, Bae EA, Han MJ, Kim DH (2005) Hepatoprotective effect of ginsenoside Rb1 and compound K on tert-butyl hydroperoxide-induced liver injury. *Liver International* 25: 1069-1073.
- Park JS, Shin JA, Jung JS, Hyun JW, Van Le, et al. (2012) Anti-inflammatory mechanism of compound K in activated microglia and its neuroprotective effect on experimental stroke in mice. *Journal Pharmacology Experimental Therapeutics* 341: 59-67.
- Su JH, Xu JH, Lu WY, Lin GQ (2006) Enzymatic transformation of ginsenoside Rg3 to Rh2 using newly isolated *Fusarium proliferatum* ECU2042. *Journal Molecular Catalysis B: Enzymatic* 38: 113-118.
- Yu H, Zhang C, Lu M, Sun F, Fu Y, et al. (2007) Purification and characterization of new special ginsenosidase hydrolyzing multi-glycosides of protopanaxadiol ginsenosides, ginsenosidase type I. *Chemical Pharmaceutical Bulletin* 55: 231-235.
- Chi H, Kim DH, Ji GE (2005) Transformation of ginsenosides Rb2 and Rc from *Panax ginseng* by food microorganisms. *Biological Pharmaceutical Bulletin* 28: 2102-2015.
- Hyun C, Lee BH, You HJ, Park MS, Ji GE (2006) Differential transformation of ginsenosides from *Panax ginseng* by lactic acid bacteria. *Journal Microbiology Biotechnology* 16: 1629-1633.
- Cheng LQ, Na JR, Kim MK, Band MH, Yang DC (2007) Microbial Conversion of Ginsenoside Rb1 to Minor Ginsenoside F2 and Gypenoside XVII by *Intrasporangium* sp. GS603 Isolated from Soil. *Journal Microbiology Biotechnology* 17: 1937-1943.
- Ko SR, Suzuki Y, Suzuki K, Choi KJ, Cho BG (2007) Marked production of ginsenosides Rd, F2, Rg3, and compound K by enzymatic method. *Chemical Pharmaceutical Bulletin* 55: 1522-1527.

17. Yan Q, Zhou W, Li X, Feng M, Zhou P (2008) Purification method improvement and characterization of a novel ginsenoside-hydrolyzing β -glucosidase from *Paecilomyces bainier* sp. 229. *Bioscience Biotechnology Biochemistry* 72: 352–359.
18. Noh KH, Son JW, Kim HJ, Oh DK (2009) Ginsenoside compound K production from ginseng root extract by a thermostable β -glucosidase from *Sulfolobus solfataricus*. *Bioscience Biotechnology Biochemistry* 73: 316–321.
19. Quan LH, Cheng LQ, Kim HB, Kim JH, Son NR et al. (2010) Bioconversion of ginsenoside Rd into compound K by *Lactobacillus pentosus* DC101 isolated from kimchi. *Journal Ginseng Research* 34: 288–295.
20. Quan LH, Min JW, Jin Y, Wang C, Kim YJ et al. (2012) Enzymatic biotransformation of ginsenoside Rb1 to compound K by recombinant β -glucosidase from *Microbacterium esteraromaticu*. *Journal Agricultural Food Chemistry* 60: 3776–3781.
21. Chang KH, Jo MN, Kim KT, Paik, H.D (2014) Evaluation of glucosidases of *Aspergillus niger* strain comparing with other glucosidases in transformation of ginsenoside Rb1 to ginsenoside Rg3. *Journal Ginseng Research* 38: 47–51.
22. Liu CY, Zhou RX, Sun CK, Jin, YH, Yu HS, et al. (2015) Preparation of minor ginsenosides C-Mc, C-Y, F2, and C-K from American ginseng PPD-ginsenoside using special ginsenosidase type-I from *Aspergillus niger* g.848. *Journal Ginseng Research* 39: 221–229.
23. Upadhyaya J, Kim MJ, Kim YH, Ko SR, Park HW, et al. (2016) Enzymatic formation of compound-K from ginsenoside Rb1 by enzyme preparation from cultured mycelia of *Armillaria mellea*. *Journal Ginseng Research* 40: 105–112.
24. Park SE, Na CS, Yoo SA, Seo SH, Son HS (2017) Biotransformation of major ginsenosides in ginsenoside model culture by lactic acid bacteria. *Journal Ginseng Research* 41: 36–42.
25. Murooka Y, Yamshita M (2008) Traditional healthful fermented products of Japan. *Journal Industrial Microbiology Biotechnology* 35: 791–798.
26. Hang YD, Woodams EE (1994) Apple pomace: A potential substrate for production of β -glucosidases from different *Aspergillus* strains. *Food Science Technology* 27: 587–589.
27. Li L, Lee SJ, Yuan QP, Im WT, Kim SC, et al. (2018) Production of bioactive ginsenoside Rg3(S) and compound K using recombinant *Lactococcus lactis*. *Journal Ginseng Research* 2018: 412–418.
28. Yan X, Fan Y, Wei W, Wang P, Liu Q, et al. (2014) Production of bioactive ginsenoside compound K in metabolically engineered yeast. *Cell Research* 24: 770–773.
29. Nan W, Zhao F, Zhang C, Ju H, Lu W (2020) Promotion of compound K production in *Saccharomyces cerevisiae* by glycerol. *Microbial Cell Factories* 19: 41.
30. Li D, Wu Y, Zhang C, Sun J, Zhou Z, et al. (2019) Production of triterpene ginsenoside compound K in the non-conventional yeast *Yarrowia lipolytica*. *Agricultural Food Chemistry* 67: 2581–2588.
31. Gwak YS, Han JY, Adhikari PB, Ahn CH (2017) Heterologous production of a ginsenoside saponin (compound K) and its precursors in transgenic tobacco impairs the vegetative and reproductive growth. *Planta* 245: 1105–1119.
32. Ratan ZA, Haidere MF, Hong YH, Park SH, Lee JO, et al. (2020) Pharmacological potential of ginseng and its major component ginsenosides. *Journal Ginseng Research*. <https://doi.org/10.1016/j.jgr.2020.02.004>
33. Kim WY, Kim JM, Han SB, Lee SK, Kim ND, et al. (2000) Steaming of ginseng at high temperature enhances biological activity. *Journal Natural Products* 63: 1702–1704.
34. Lim CY, Moon JM, Kim BY, Lim SH, Lee GS, et al. (2015) Comparative study of Korean white ginseng and Korean red ginseng on efficacies of OVA-induced asthma model in mice. *Journal Ginseng Research* 39: 38–45.